

Serologic Abnormalities in Autoimmune Pancreatitis

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Introduction

In 1961, Sarles et al reported a case of pancreatitis with hypergammaglobulinemia, which in retrospect appears to be identical to autoimmune pancreatitis (AIP) (26). In 1995, Yoshida et al. described such a case as AIP (37). In 2001, Hamano et al. later reported increased serum levels of IgG4 in AIP (9). The histopathological findings of AIP are characterized by the periductal localization of predominantly CD4 positive T-cells, IgG4-positive plasma cells, storiform fibrosis with acinar cell atrophy frequently resulting in the stenosis of the main pancreatic duct, and obliterative fibrosis, resulting in the so called lymphoplasmacytic sclerosing pancreatitis (LPSP) (13, 22, 23, 27). Although the infiltration of IgG4-positive cells and increased serum levels of IgG4 are characteristic, they are not specific for type 1 AIP and the role of IgG4 in the development of AIP and IgG4-related disease remains unclear (10, 34, 35).

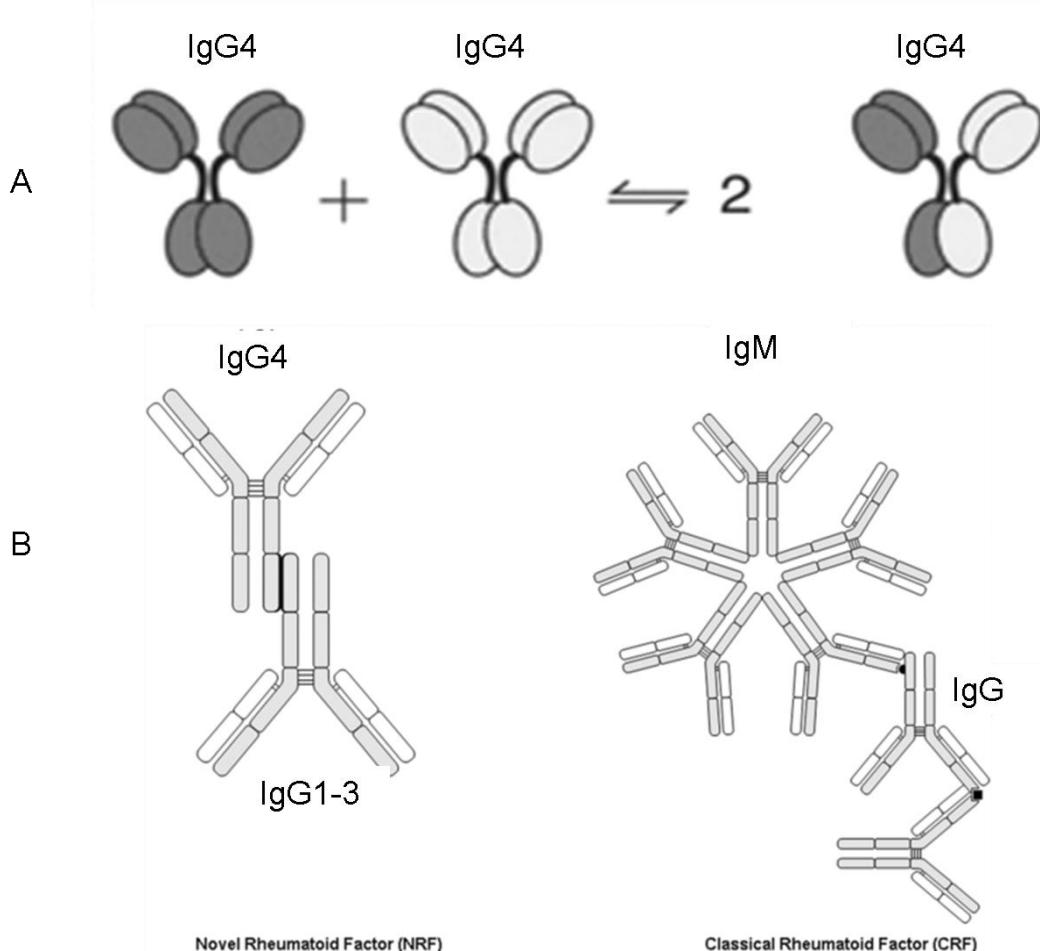
Immunology of immunoglobulin subclasses

Generally, the amount of IgG4 does not vary with sex or age, and the quantity of IgG4 as well as the IgG4/total IgG ratio tends to remain constant (25). In normal subjects, IgG4 consists of 4–6% of total IgG, and its serum elevation has been seen in several conditions, such as allergic disease, parasite infection, and pemphigus vulgaris (25). In type 1 AIP, total IgG, IgG1, IgG2, IgG4 and IgE were usually increased compared with healthy subjects, while IgM, IgA, and the ratios of IgG to IgM or IgA, are decreased compared with normal or other control diseases

(9, 28) (**Table 1**). In AIP, all subclasses of IgG were increased compared with other types of pancreatitis.

Although the association with IgE-mediated allergy and IgG4 antibodies is well-known, the characteristics of IgG4 are less understood (24). IgG4 antibodies participate in a continuous process referred to as Fab-arm exchange, which describes swapping a heavy chain and attached light chain (half-molecule) with a heavy-light chain pair from another molecule (36). This results in asymmetric antibodies with two different antigen-combining sites. While these modified antibodies are hetero-bivalent, they behave as monovalent antibodies (**Figure 1A**) (36). Another aspect of IgG4 mimics IgG rheumatoid factor (RF) activity by interacting with IgG on a solid support (**Figure 1B**) (11). In contrast to conventional RF, which binds via its variable domains, the activity of IgG4 is located in its constant domains, but is inefficient to activate potentially dangerous effector systems due to its low affinity for C1q and the classical Fc_y-receptors.

Comparison of various markers in differentiating between AIP and pancreatic cancer showed that the best results are obtained using IgG4, which has 86% sensitivity, 96% specificity, and 91% accuracy (**Table 2**) (12). IgG4 was therefore adopted as the best marker in the diagnostic criteria of type 1 AIP (12). However, serum IgG4 elevation or numerous IgG4-bearing plasma cell infiltrations has been reported in some patients with pancreatic cancer, suggesting that these features are not completely specific for AIP and cannot exclude the presence of pancreatic cancer (7).



Characteristic forms of IgG4.

1a. Schematic representation of the generation of bispecific IgG4 antibodies by the exchange of half-molecules ('Fab-arm exchange') (cited from (36) with permission). IgG4 Fab arm exchange occurs by the exchange of a heavy chain–light chain pair (half-molecule) of one IgG4 molecule with that of another IgG4 molecule. The IgG4 molecule may thereby acquire two distinct Fab arms and become bispecific. The Fc structure remains essentially unchanged apart from potential changes due to differences in glycosylation or allotype. Fab arm exchange is proposed to be stochastic and dynamic.

1b. On the left: IgG4 Fc interacts with Ig Fc. On the right: IgM RF recognizes IgG in a "classical" Fab-Fc recognition (cited from (11) with permission). **Table 1.** Serum immunoglobulin levels in patients with autoimmune pancreatitis (9, 28). *Data from reference (25). AIP, autoimmune pancreatitis; CP, chronic pancreatitis; IC, immune complex.

Table 1. Serum immunoglobulin levels in patients with autoimmune pancreatitis (9, 28).

	Year	N	IgG	IgG1	IgG2	IgG3	IgG4/IgG	IgM	IgA	IgE	IC(ug/ml)
Hamano et al	2001	AIP	20	2,201	868	617	53	663 (30 %)	91	226	176
		Control	20	1,341	664	592	34	51		142	247
Taguchi et al	2009	AIP	20	2,556	NT	NT	NT	762	85	213	NT
		CP	21	1,245*	NT	NT	NT	NT	122	294	NT

*Data from reference (25). AIP, autoimmune pancreatitis; CP, chronic pancreatitis; IC, immune complex.

Table 2. Comparison of various markers in the differentiation of autoimmune pancreatitis and pancreatic cancer using identical sera.

	Sensitivity (AIP n = 100) (%)	Specificity (vs. PC n = 80) (%)	Accuracy (vs. PC)
IgG4	86	96	91
IgG	69	75	72
ANA (anti-nuclear antibody)	58	79	67
RF (rheumatoid factor)	23	94	54
IgG4+ANA	95	76	87
IgG+ANA	85	63	75
IgG4+IgG+ANA	95	63	81
IgG4+RF	90	90	90
IgG+RF	78	73	76
IgG4+IgG+RF	91	71	82
ANA+RF	69	60	78
IgG4+ANA+RF	97	73	86
IgG+ANA+RF	91	61	78
IgG4+IgG+ANA+RF	97	61	81

AIP autoimmune pancreatitis, PC pancreatic cancer

(Cited from reference (12) with permission)

The complement system

Patients with active AIP occasionally have decreased complement (C3, C4) levels with elevated circulating immune complexes and serum IgG4 elevation (4, 9). However, a recent study showed that the classical pathway of complement activation through IgG1 may be involved in the development of AIP rather than mannose-binding lectin or alternative pathways through IgG4 (16). Moreover, IgG4 bound to other isotypes such as IgG1, IgG2, and IgG3 with an Fc-Fc interaction develop immune complexes in patients with AIP. In this setting IgG4 may contribute to the clearance of immune complexes or termination of the inflammatory process by preventing the formation of large immune complexes by blocking the Fc mediated effector functions of IgG1 (11). Compared with SLE, tubulointerstitial nephritis (TIN) is more often observed in renal lesions of IgG4-related disease. In TIN associated with AIP deposition of immune complex (IgG and C3) is more commonly observed in the tubular basement membrane, rather than the glomerular basement membrane as is typically seen in SLE (32).

Autoantibodies

In addition to increased total IgG and IgG4, patients with IgG4-related disease often have detectable autoantibodies, albeit not organ-specific (22,23). Although some patients with IgG4-related disease have non-specific antibodies such as anti-nuclear antibody (ANA), there is no clear association, aside from overlapping symptoms, between IgG4-related disease and common autoimmune diseases such as Sjögren's syndrome and SLE. In regards to IgG4 function, it remains unclear if IgG4-related disease is a true autoimmune or allergic disease. However, the frequent coexistence of other organ involvement lead to the concept that there may be common target antigens in the involved organs such as the pancreas, salivary gland, biliary tract, lung, renal tubules, etc.. Although the disease specific antibodies have not been identified several disease-related antibodies such as anti-lactoferrin (LF) (20,30), anti-carbonic anhydrase (CA)-II (2, 17, 20, 30), anti-CA-IV (19), anti-pancreatic secretory trypsin inhibitor (PSTI) (3), anti-amylase-alpha (5), anti-HSP-10 (29), and anti-plasminogen-binding protein (PBP) peptide autoantibodies (6) have been reported. Although patients have increased serum levels of IgG4, the major subclass of these autoantibodies is not necessarily IgG4, but often IgG1 (3). CA-II (20), CA-IV (19), LF (20) and PSTI (3) are distributed in the ductal cells of several exocrine organs, including the pancreas, salivary gland, biliary duct, and lung (17,20). Although all peptides have not been systematically studied, immunization with CA-II or LF induced systemic lesions such as pancreatitis, sialadenitis, cholangitis, and interstitial nephritis in mice models appear similar to human IgG4-related diseases (18, 33). The high prevalence of these antibodies suggests that they are at least potential candidate target antigens in AIP (20, 30).

Molecular mimicry among microbes and target antigens may be a possible mechanism to overcome immune tolerance. This hypothesis is based on the concept that infectious agents

share one or more epitopes with self-components, or infectious agents cause bystander activation of immune cells with autoaggressive potential (8,14,15). Guarneri and colleagues showed significant homology between human CA-II and alpha-CA of *H. pylori*, a fundamental enzyme for bacterial survival and proliferation in the stomach (8). Moreover, the homologous segments contain the binding motif of DRB1*0405, which confers a risk for AIP development (8). The PBP peptide identified in European patients with AIP shows homology with an amino acid sequence of PBP of *H. pylori* and with ubiquitin-protein ligase E3 component n-recognition 2 (UBR2), an enzyme highly expressed in acinar cells of the pancreas (6). These findings suggest that gastric *H. pylori* infection might trigger AIP in genetically predisposed subjects (8, 14, 15).

Diabetes mellitus is affects 43~68% of the patients with AIP, but autoantibodies against

glutamic acid decarboxylase, beta-cell or tyrosine phosphatase-like protein associated type1 DM are rarely observed(21). These findings suggest that islet cells are not likely targeted in the development of DM associated with AIP.

Summary

Although serum IgG4 elevation is a characteristic finding and useful to establish a diagnosis of type 1 AIP, it is not specific for this disorder. Importantly it can be seen in other conditions with similar clinical presentations, including pancreatic cancer. The role of IgG4 antibodies in the pathogenesis of AIP and IgG4-related disease remains unclear. Several autoantibodies have been identified in subjects with AIP, but additional studies are needed to clarify the significance of these findings in the pathophysiology of AIP.

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