



Genetics of acute pancreatitis

F. Ulrich Weiss and Markus M. Lerch Department of Medicine A, University Medicine Greifswald, Ferdinand-Sauerbruch-Strasse, 17489 Greifswald, Germany e-mail: ulrich.weiss@uni-greifswald.de

Version 1.0, August 17, 2016 [DOI: 10.3998/panc.2016.22]

1. Introduction

It becomes increasingly apparent that acute pancreatitis (AP), recurrent acute pancreatitis (RAP) and chronic pancreatitis (CP) represent overlapping phenotypes of a single disease entity and the latter may begin in the guise of the former. Accordingly the genetic risk factors that have been identified for chronic pancreatitis have also been found to be of some relevance in acute and recurrent acute pancreatitis. Other inherited factors have been found to not influence disease susceptibility but rather the disease severity of AP. The most prominent in this category are genes that regulate cytokines and inflammatory response proteins. This chapter reviews the genetic changes that confer disease susceptibility as well as severity in patients with acute pancreatitis.

2. Definition and Diagnosis

Acute pancreatitis is a syndrome of a sudden pancreatic inflammation with unpredictable severity, duration, complications and outcome.

The diagnosis of acute pancreatitis requires two of the following three features: 1) *abdominal pain* consistent with acute pancreatitis (acute onset of a persistent, severe, epigastric pain often radiating to the back); 2) serum *lipase activity (or amylase activity) at least three times greater than the upper limit of normal*; and 3) characteristic findings of acute pancreatitis on contrastenhanced computed tomography (CECT) and less commonly magnetic resonance imaging (MRI) or transabdominal ultrasonography (9).

The main etiologic causes of acute pancreatitis are gallstones and alcohol abuse but other rare causes include trauma, endoscopic interventions, infections and drugs or toxins. Also conditions such as hypercalcemia or hypertriglyceridemia have been suggested to increase the risk for acute pancreatitis. The onset of a "first" AP episode is caused by an acute injury of pancreatic tissue that rapidly disrupts its normal physiologic function and initiates an acute inflammatory response. Histologic damage occurs as a consequence of intra-acinar activation of digestive enzymes and a subsequent infiltration of pancreatic tissue with inflammatory cells. This pro-inflammatory cascade is normally self-limited and followed by anti-inflammatory responses which may include the activation of pancreatic stellate cells and the start of fibrosis. Clinical recovery from AP episodes is attained in most cases within 3-5 days. The etiology of acute pancreatitis can be established in approximately 75 percent of patients, leaving one in four patients with so called idiopathic acute pancreatitis. In to AP а consequence contrast as of environmental factors, inherited forms present with an earlier onset of acute or recurrent acute pancreatitis that eventually will progress to chronic disease. Here we discuss genetic mutations that have been associated with acute pancreatitis and influence the phenotype of the disease.

3. Acute, recurrent acute and chronic pancreatitis.

CP is a progressive inflammatory disease that may develop from acute to recurrent and chronic disease states. Historically, chronic pancreatitis has been associated with alcoholism and many CP patients are suspected of alcohol abuse, often unjustly (3). Today growing evidence suggests that genetic risk factors contribute substantially to the pancreatitis risk in CP patients (35, 43). In contrast to sporadic attacks of gallstone or alcohol-induced pancreatitis, hereditary forms of pancreatitis (HP) typically present in childhood with attacks of acute pancreatitis (AP) that become recurrent. Over time, HP patients with recurrent episodes of pancreatitis in the absence of precipitating factors may develop the same common complications as alcoholic CP patients, pancreatic fibrosis, pseudocyst including formation, pancreatic exocrine insufficiency (PEI) and diabetes mellitus. Large cohort studies on pancreatitis have established complex interactions between multiple genetic and environmental factors in the progression from RAP to CP. Clinical implications of genetic risk factors have not been established due to prognostic or therapeutic limitations of current genetic testing modalities.

4. Acute pancreatitis in children

There have been several studies in the last years reporting increasing incidences of acute pancreatitis among pediatric patients (49, 55, 59, 64). Current estimates range from 3.6-13.2 cases per 100,000 children, which is close to the incidence of AP in adults (59). Underlying causes may involve increased testing of amylase and lipase serum levels, more frequent emergency department visits and improved clinical awareness. The rising incidence of obesity in children may also contribute as an independent risk factor for acute biliary pancreatitis, which was once not a common etiology among children (44). A recent national survey of 55,000 hospitalized children (1-20 years old) with acute pancreatitis in the United States revealed that AP occurs more frequently in children older than 5 years (62.8% were older than 15 years) and slightly more frequently in girls (63%) than in boys (58). Hepatobiliary disease was the comorbidity condition with the greatest association with AP in this study, whereas other studies claim that the change in incidence of AP is primarily due to an increase of cases with systemic diseases and those with an unidentified (idiopathic) etiology (55).

Considerable differences exist in the etiology of AP in adults and children. Whereas 70% of cases in adults can be attributed to the gallstones or alcohol abuse, the causes of AP in children are more diverse. In a recent study by Bai et al. (7) the top 5 etiologies of acute pancreatitis in children were biliary, medications, idiopathic, systemic disease, and trauma, followed by infectious, metabolic, and hereditary causes. Not surprisingly, alcohol was not reported as a common cause of pancreatitis in children and genetic mutations were identified in about 5-8% of patients. Mutations were most commonly found in the cationic trypsinogen gene (PRSS1), the pancreatic secretory trypsin inhibitor gene (SPINK1), and the cystic fibrosis transmembrane conductance regulator gene (CFTR). A hereditary etiology is nearly indistinguishable clinically or by imaging from other causes of AP. Early onset, and recurrent events occurring during the first decade of life in combination with a family history may be the best indication for a genetic background of AP.

In a retrospective genetic analysis of 69 children with recurrent acute pancreatitis (RAP) or CP, Vue et al. identified 48% as a carrier of at least 1 mutation in one of the known risk genes of chronic pancreatitis: *PRSS1, CFTR* and *SPINK1* (73). Patients with mutations were more likely to have a family history but otherwise could not be identified by any mutation-specific phenotypic differences. Similar results were obtained by Palermo et al. in a genetic analysis of 45 pediatric AP patients, of which 60% carried a least one mutation in one of the pancreatitis risk genes PRSS1, SPINK1, CTRC and CFTR. Even though the study cohort has not been completely genotyped, the authors claim that they identified a higher frequency of CFTR mutations in CP patients in comparison to RAP patients. A multinational cross-sectional study of 301 children with RAP and CP has been performed by the INSPPIRE consortium (40). 84% of children with CP reported prior recurrent episodes of AP. Sequencing analysis identified at least one mutation in pancreatitis-related genes in 48% of patients with RAP vs 73% of patients with CP. Children with PRSS1 or SPINK1 mutations were more likely to develop CP, but also ethnic differences seem to affect the disease phenotype and disease progression. A higher disease burden in CP patients might justify early genetic testing in pediatric AP patients which may also help to optimize therapeutic strategies to stop disease progression in these patients

5. Risk genes

Two decades of worldwide screening efforts have confirmed а complex network of aeneenvironment interactions that control or influence the development and progression of pancreatic diseases including acute, recurrent acute and chronic pancreatitis. While acute pancreatitis in most cases can be attributed to environmental factors such as gallstones or alcohol abuse, in 20-25% the etiology remains unclear. In these idiopathic AP patients genetic risk factors are found to play a major role in the onset of the disease. Most CP patients report prior episodes of AP or RAP and also hereditary chronic pancreatitis starts in most mutation carriers with a first attack of acute pancreatitis. The known genetic risk factors of CP therefore also play a role in the onset of acute and recurrent acute pancreatitis episodes and are identified in genetic association studies of AP patients, however at a lower incidence rate when compared to patients diagnosed with idiopatic CP. Most identified genetic risk factors to date are involved in the regulation and control of protease activity, starting with the initial identification of an autosomal dominant mutation in the cationic trypsinogen (PRSS1) in 1996 by Whitcomb (75). Candidategene approaches and validation studies in multiple cohorts have increased the number of pancreatitis-related risk genes, which include CFTR, SPINK1 and CTRC (16, 66, 69, 78, 79). Significant association with pancreatitis has also been demonstrated for sequence variants in CPA1, CASR, and CEL (29, 50, 78). Preliminary reports which await further validation include CLDN2 (76), CTSB (46), MYO9B (54) and UBR1 (83) or the association of an increased pancreatitis risk with ABO blood group and the socalled "secretor status", which is determined by a mutation of the Fucosyl-transferase gene FUT2 (74). With the exception of the dominant PRSS1 mutations, most variant alleles of these risk genes are not single factor causes, but predisposing to pancreatitis and may lower the threshold for pancreatitis attacks. They predispose not only to acute pancreatitis, but to recurrent episodes or the progression to chronic disease. Additional environmental or metabolic factors are operative and relevant in the complex gene-environmental interactions that determine the disease phenotype in each individual patient.

Metabolic causes of pancreatitis are less common, but also constitute an important component of the etiologic factors of AP. They include hypercalcemia, hypertriglyceridemia, diabetes mellitus and rarely Wilson's disease (39). Familial Hypocalciuric Hypercalcemia (FHH) was first described in the 1970s (30), which led to the subsequent cloning of the calcium-sensing receptor (CASR) and the discovery of its pivotal role in disorders of calcium homeostasis like FFH (61). The CaSR plays a central role in calcium homeostasis primarily by regulation of parathyroid hormone (PTH) secretion and calcium reabsorption in the renal tubular system. Pearce et al. reported in 1996 three FHH kindreds with recurrent pancreatitis and in all patients the disease was associated with missense mutations in the extracellular domain of the CaSR (60). In acinar cells low calcium concentrations are prevalent in the cytosol, which constitutes one failsafe mechanism in the prevention of intra-acinar activation. Hypercalcemia-related trypsinogen pancreatitis can also be secondary to primary hyperparathyroidism (PHPT) and was first reported by Cope et al. in 1957 (17). PHPT represents a non-physiological overproduction of parathyroid hormone, caused by adenoma of the parathyroid gland or multiple endocrine neoplasia (MEN) types 1 and 2A. Genetic studies provide evidence that inherited mutations in pancreatitisrelated genes SPINK1 and CFTR, but not CASR were identified in 36% of hyperparathyroidism patients who developed AP (26, 27, 42). A recent review of the literature by Bai et al. (6) confirmed an association of PHPT with pancreatitis and implicates hypercalcemia, but the functional role of CaSR mutations in the context of pancreatitis has yet to be elucidated. Apparently PHTP requires multiple genetic and environmental influences to induce pancreatitis.

Another minor but significant etiologic factor of acute pancreatitis are familial disorders, including lipoprotein lipase deficiency, apolipoprotein C-II deficiency or common hypertriglyceridemia that lead to plasma accumulations of chylomicrons or triglycerides. Lipoprotein lipase catalyzes the hydrolysis of triglyceride from chylomicrons and very low density lipoprotein (VLDL) and therefore plays a central role in the regulation of the energy metabolism. Familial lipoprotein lipase deficiency prevents the enzyme from effectively breaking down triglycerides in the bloodstream and leads to chylomicronemia and consequently very severe hypertriglyceridemia. As a result, triglycerides attached to lipoproteins accumulate in plasma and tissues, leading to inflammation of the pancreas enlarged (pancreatitis), liver and spleen (hepatosplenomegaly), and fatty deposits in the skin (eruptive xanthomas). Triglyceride levels above 2,000 mg/dl should be considered a significant risk factor of developing pancreatitis (12).

The most common familial disorders associated with chylomicronemia are the type I and type V hyperlipoproteinemias (10). Hyperlipoproteinemia type I is caused by loss-of function mutations in the LPL gene or in the gene of its co-factor ApoC2 (18) and is inherited in an autosomal recessive pattern. The frequency of LPL deficiency in the general population is estimated to be about 1-2 per million (13). More than 100 LPL sequence variants have been described, most of them associated with a loss of catalytic activity (48, 63). LPL deficient patients are homozygote or compound heterozygote for these mutations and work on a systematic classification of LPL gene variants is ongoing (65). Also, rare mutations in like other genes the apoA5. glycosylphosphatidylinositol-anchored hiahdensity lipoprotein-binding protein 1 (GPIHBP1) or lipase maturation factor 1 (LMF1) have been reported to affect LPL acitivity and were found to associate with chylomicronemia (1, 38).

Plasma triglyceride levels are also elevated as a result of hepatic over-production of VLDL or heterozygous LPL deficiency in familial hypertriglyceridemia Type IV. This monogenic familial hypertriglyceridemia is associated with only mild hypertriglyceridemia. Additional increases of plasma lipid levels in these predisposed patients may arise from unrelated risk factors like plasmocytoma, systemic lupus erythematosus and lymphomatous disease and enhance the risk further of developing pancreatitis.

Another very rare autosomal recessive metabolic disorder with associated acute pancreatitis is the congenital lipodystrophy, or Berardinelli-Seip congenital lipodystrophy (BSCL) [OMIM 269700], which has an estimated prevalence of one in 10 million. Affected patients have a generalized muscular appearance due to the nearly complete absence of fat tissue (11) and present with tryglyceridemia, hepatomegaly, mental retardation, insulin-resistant diabetes mellitus and hypertrophic cardiomyopathy (32). Hypertriglyceridemia seems the to be

predisposing factor for the development of acute pancreatitis, however the pathophysiology and genetic background of the disease have not been completely resolved. Linkage analysis identified mutations in the 1-acylglycerol-3-phosphateOacetyltransferase 2 (AGPAT2), a key enzyme in biosynthesis triacyglycerol the of and alycerophospholipids (2) and in a second locus, BSCL2/seipin at 11q13 (45), with sequence homology to a murine guanine nucleotide-binding protein γ 3-linked gene (Gng3lg). BSCL patients are homozygous or compound heterozygous carriers of loss-of function mutations.

6. Disease Severity and Prognostic Marker

Acute pancreatitis has an annual incidence of 10-30 per 100,000 population (4, 23). 80% of AP episodes have a mild course, without significant morbidity or mortality. However, in 20% of cases the disease is severe with a mortality rate of 25% to 30%.(41, 52) Increased serum levels of TNF- α , IL-1, IL-6, IL-8 and their (soluble) receptors in AP patients demonstrate an important role of these major early cytokines (14, 20) which mediate the systemic inflammatory response. Interleukin secretion is regulated at the transcriptional level which makes SNPs in the promoter region of these inflammatory mediators likely risk factor candidates for the development of systemic inflammatory response syndrome (SIRS) and organ failure. Variants TNF- α -238G>A, and -308G>A have been identified as transcriptional enhancers, leading to higher *TNF*- α levels (8, 77). In a recent meta-analysis on more than 1500 patients and 1330 controls from 12 published case-control studies Yang et al demonstrated that the common *TNF*- α polymorphisms (-238, -308) do not alter the risk of pancreatitis nor have they any influence on disease severity (shown only for the -308 SNP) (80).

Also some polymorphisms in the promoter regions of IL-1 β , (-511C>T, -31C>T, +3954C>T), IL-6 (-634C>G, -174G>C), IL-8 (-251T>A), and IL-10 (-

1082A>G, -819C>T, -592C>A) were identified to affect transcriptional activities and therefore were considered as potential risk factors for disease severity (24, 28, 51, 57). The second intron of the IL1Ra gene (IL1RN) contains in addition a variable number of tandem repeats (VNTR) of 86 nucleotides and carriers of allele 2, containing 2 repeats, face increased production of IL1Ra protein. Some genetic association studies have suggested that different IL1RN alleles are associated with specific disease risks for sepsis (25), ulcerative colitis (47) or increase the susceptibility to gastric cancer (85). A number of aenetic association studies limited have investigated these polymorphisms in different population cohorts of acute pancreatitis patients, but showed inconclusive results (19, 62, 72). The IL-1 gene cluster had been implicated in acute pancreatitis by a study of Smithies et al in 2000, but they found no association of the IL-1β+3954C>T polymorphism in a cohort of British AP patients (70). Also the IL-10 -1082A>G, -819C>T and -592C>A polymorphisms did not associate with AP among British AP patients (67). In contrast, Hofner reported a significant association of the IL-8 -251T>A polymorphism with the risk for AP (34). In a recent meta-analysis in 2013 Yin and colleagues evaluated 10 studies on IL gene polymorphisms and their susceptibility to AP (81). Their study suggests that indeed the IL-8 -251T>A polymorphism is associated with an increased risk of AP. However, no risk association could be confirmed for any of the gene polymorphisms in IL-1B, IL-6 or IL-10.

IL-1 constitutes actually a group of cytokines produced by a wide range of cells including macrophages, monocytes, fibroblasts and dendritic cells and elicits the acute phase response of the body against infection. IL-1α and IL-1β are the most analyzed members which have a natural antagonist (IL1Ra) and they all bind to the same type I IL-1 receptor (IL-1RI). Polymorphisms in IL-1 genes have been found associated with some cancers and Grave's disease (22). In a meta-analysis of 37 studies Ying recently reported that IL-1β-31C>T polymorphism might confer susceptibility to gastric cancer in the presence of H.pylori infection, indicating gene-environment interaction in gastric carcinogenesis (82). Another analysis of 11 case-control studies by Chen et al. confirmed significant protection of the IL-1b-511C>T polymorphism from Grave's Disease in Asians, but not in Caucasians (15). These results may indicate that the pancreatitis risk evaluation of IL polymorphisms is more complex than previously thought and future studies should be carefully genetic consider background designed to genedifferences. as well as additional environment interactions.

The interleukin 1 family is also closely linked to the innate immune response and the cytoplasmic region of the type I IL-1 receptor is highly homologous to the cytoplasmic domains of the Toll-like receptors (TLRs).

TLRs play a critical role in the development of pancreatic diseases as they mediate interactions between environmental stimuli and the innate immune response. They belong to a larger family of so-called pattern-recognition receptors (PRRs) which are activated by either pathogen-associated or damage-associated molecular patterns (PAMPs or DAMPs) (37), released by activated or necrotic cells in response to stress or cell involve TLR signaling damage. Myeloid differentiation primary response protein (MyD88)pathways upregulate dependent and the transcription of pro-inflammatory genes through the activation of NFkB (36). TLR3 and TLR4 can further activate in addition the TRIF pathway leading to the synthesis of interferon- α/β (56).

Sequence variations in TLR genes are capable of influencing the susceptibility to infectious diseases (53, 68) and the common TLR4 polymorphisms p.D299G and p.T399I were the first identified risk factors for the development of sepsis in patients (5). Variant TLR4 receptors show reduced interaction with lipopolysaccharide which may result in higher infection with Gram-negative bacteria. A first limited genetic association study by Hofner (34) did not find an association of these TLR4 polymorphisms with the incidence or severity of acute pancreatitis, but several subsequent studies in Caucasian and Asian patient cohorts had inconsistent results (31, 33, 71, 84). A meta-analysis by Zhou on 1255 cases and 998 controls did not confirm a risk factor role of TLR4 D299G and T399I polymorphisms for AP susceptibility (86). A number of additional genetic analyses have been performed, which reported significant association of TLR2 intronic mutation with susceptibility and severity of AP in Japan (71), and a risk factor role of Mannose-binding lectin (MBL) promoter variants with disease severity in Chinese patients (84). These studies await confirmation.

Also, the role of pattern-recognition receptors in the pathophysiology of mucosal barrier failure in acute pancreatitis has currently not been resolved. A disturbance of the mucosal barrier plays an essential role in the development of severe acute pancreatitis, as it allows bacterial translocation from the gut into the blood stream, which may trigger infectious complications. Nijmejier et al. performed a candidate gene approach in more than 500 AP patients from the Netherlands and Germany and reported that sequence variants in myosin IXB (MYO9B), a protein which seems to play a role in tight junction assembly, do not only associate with inflammatory bowel disease (IBD) and celiac disease (CD), but also with acute pancreatitis (54). Myosin IXB variants may confer a higher risk of intestinal barrier dysfunction in AP.

7. Identification of additional Risk factors

After the publication of the first pancreatitisassociated risk gene in 1996 by Whitcomb et al. the identification of genetic risk factors in pancreatitis followed for two decades mainly candidate gene approaches. These efforts were successful and contributed significantly to our current understanding of the molecular details of pancreatic pathophysiology. Powerful new screening technologies include Genome-wide association analyses and Next Generation Sequencing studies. These techniques are rather expensive and require large cohorts of clinically well-defined individuals but they are ideally suited to identify new risk factors outside the already known or suspected signaling pathways or regulatory mechanisms involved in the pancreas physiology. To date few GWA studies have been performed, mainly in CP and RAP patients, and the study by Whitcomb et al. was able to identify one new susceptibility locus in the Claudin-2 gene (CLDN2) (76). A second SNP found in the PRSS1-PRSS2 locus seems to further confirm the importance of this established risk locus for the development of pancreatitis. In a replication GWA study on European patients with alcoholic and non-alcoholic CP, Derikx and colleagues were able to confirm these findings (21). A third GWA was done by Weiss et al. on high serum lipase values in a population-based cohort of healthy individuals (74). The study reported an association of blood group B (ABO-B) and the non-secretor allele of the fucosyltransferase-2 locus (FUT2) with elevated lipase activities in asymptomatic individuals. Both loci were also identified to associate with CP. Also this study awaits confirmation in larger replication cohorts involving different ethnic populations.

These new findings and upcoming reports from candidate-free current genetic screening approaches may open the route for studies on pathological mechanisms outside the known protease-antiprotease homeostasis network. More GWA studies and NGS-data will significantly contribute to expand our current understanding of pancreatic pathophysiology and pancreatic disease and hopefully will help to identify new therapeutic strategies.

8. References

- 1. Adeyo O, Goulbourne CN, Bensadoun A, Beigneux AP, Fong LG and Young SG. Glycosylphosphatidylinositol-anchored high-density lipoprotein-binding protein 1 and the intravascular processing of triglyceride-rich lipoproteins. *J Intern Med* 272(6): 528-540, 2012. PMID: 23020258.
- Agarwal AK, Arioglu E, De Almeida S, Akkoc N, Taylor SI, Bowcock AM, et al. AGPAT2 is mutated in congenital generalized lipodystrophy linked to chromosome 9q34. Nat Genet 31(1): 21-23, 2002. <u>PMID:</u> 11967537.
- 3. Aghdassi AA,Weiss FU,Mayerle J,Lerch MM and Simon P. Genetic susceptibility factors for alcoholinduced chronic pancreatitis. *Pancreatology* 15(4 Suppl): S23-31,2015. PMID: 26149858.
- 4. Andersson R, Andersson B, Haraldsen P, Drewsen G and Eckerwall G. Incidence, management and recurrence rate of acute pancreatitis. *Scand J Gastroenterol* 39(9): 891-894, 2004. <u>PMID: 15513389.</u>
- 5. Arbour NC, Lorenz E, Schutte BC, Zabner J, Kline JN, Jones M, et al. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet* 25(2): 187-191, 2000. PMID: 10835634.
- 6. Bai HX, Giefer M, Patel M, Orabi Al and Husain SZ. The association of primary hyperparathyroidism with pancreatitis. *J Clin Gastroenterol* 46(8): 656-661, 2012. <u>PMID: 22874807.</u>
- 7. Bai HX, Lowe ME and Husain SZ. What have we learned about acute pancreatitis in children? J Pediatr Gastroenterol Nutr 52(3): 262-270. PMID: 21336157.
- Balog A, Gyulai Z, Boros LG, Farkas G, Takacs T, Lonovics J, et al. Polymorphism of the TNF-alpha, HSP70-2, and CD14 genes increases susceptibility to severe acute pancreatitis. *Pancreas* 30(2): e46-50, 2005. <u>PMID: 15714129.</u>
- Banks PA, Bollen TL, Dervenis C, Gooszen HG, Johnson CD, Sarr MG, et al. Classification of acute pancreatitis-2012: revision of the Atlanta classification and definitions by international consensus. *Gut* 62(1): 102-111, 2012. <u>PMID: 23100216.</u>
- 10. Beaumont JL, Carlson LA, Cooper GR, Fejfar Z, Fredrickson DS and Strasser T. Classification of hyperlipidaemias and hyperlipoproteinaemias. *Bull World Health Organ* 43(6): 891-915, 1970. <u>PMID:</u> 4930042.
- 11. Berardinelli W. An undiagnosed endocrinometabolic syndrome: report of 2 cases. *J Clin Endocrinol Metab* 14(2): 193-204, 1954. <u>PMID: 13130666.</u>

- Berglund L, Brunzell JD, Goldberg AC, Goldberg IJ, Sacks F, Murad MH, et al. Evaluation and treatment of hypertriglyceridemia: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab 97(9): 2969-2989, 2012. <u>PMID: 22962670.</u>
- Brunzell JDaD, S.S. Familial lipoprotein lipase deficiency, apo CII deficiency and hepatic lipase deficiency. The Metabolic and Molecular Basis of Inherited Disease. Sciever C, Beaudet A, Sly WS, Vale D., New York McGraw-Hill 2789-2816, 2001.
- 14. Chen CC, Wang SS, Lee FY, Chang FY and Lee SD. Proinflammatory cytokines in early assessment of the prognosis of acute pancreatitis. *Am J Gastroenterol* 94(1): 213-218, 1999. <u>PMID: 9934758</u>.
- Chen ML, Liao N, Zhao H, Huang J and Xie ZF. Association between the IL1B (-511), IL1B (+3954), IL1RN (VNTR) polymorphisms and Graves' disease risk: a meta-analysis of 11 case-control studies. *PLoS One* 9(1): e86077, 2014. <u>PMID: 24465880.</u>
- 16. Cohn JA, Friedman KJ, Noone PG, Knowles MR, Silverman LM and Jowell PS. Relation between mutations of the cystic fibrosis gene and idiopathic pancreatitis. *N Engl J Med* 339(10): 653-658, 1998. <u>PMID:</u> 9725922.
- 17. Cope O, Culver PJ, Mixter CG, Jr. and Nardi GL. Pancreatitis, a diagnostic clue to hyperparathyroidism. Ann Surg 145(6): 857-863, 1957. <u>PMID: 13425295.</u>
- 18. Cox DW, Wills DE, Quan F and Ray PN. A deletion of one nucleotide results in functional deficiency of apolipoprotein CII (apo CII Toronto). *J Med Genet* 25(10): 649-652, 1988. PMID: 3225819.
- de-Madaria E, Martinez J, Sempere L, Lozano B, Sanchez-Paya J, Uceda F, et al. Cytokine genotypes in acute pancreatitis: association with etiology, severity, and cytokine levels in blood. *Pancreas* 37(3): 295-301, 2008. PMID: 18815552.
- 20. de Beaux AC, Goldie AS, Ross JA, Carter DC and Fearon KC. Serum concentrations of inflammatory mediators related to organ failure in patients with acute pancreatitis. *Br J Surg* 83(3): 349-353, 1996. <u>PMID:</u> 8665189.
- 21. Derikx MH, Kovacs P, Scholz M, Masson E, Chen JM, Ruffert C, et al. Polymorphisms at PRSS1-PRSS2 and CLDN2-MORC4 loci associate with alcoholic and non-alcoholic chronic pancreatitis in a European replication study. *Gut* 64(9): 1426-1433, 2014. <u>PMID: 25253127.</u>
- 22. Eland IA, Sturkenboom MJ, Wilson JH and Stricker BH. Incidence and mortality of acute pancreatitis between 1985 and 1995. Scand J Gastroenterol 35(10): 1110-1116, 2000. PMID: 11099067.
- 23. EI-Omar EM, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, et al. Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature* 404(6776): 398-402, 2000. PMID: 10746728.
- 24. Eskdale J, Gallagher G, Verweij CL, Keijsers V, Westendorp RG and Huizinga TW. Interleukin 10 secretion in relation to human IL-10 locus haplotypes. *Proc Natl Acad Sci U S A* 95(16): 9465-9470, 1998. PMID: 9689103.
- 25. Fang F, Pan J, Li Y, Xu L, Su G, Li G, et al. Association between interleukin 1 receptor antagonist gene 86bp VNTR polymorphism and sepsis: a meta-analysis. *Hum Immunol* 76(1): 1-5, 2014. <u>PMID: 25500257.</u>
- 26. Felderbauer P, Karakas E, Fendrich V, Bulut K, Horn T, Lebert R, et al. Pancreatitis risk in primary hyperparathyroidism: relation to mutations in the SPINK1 trypsin inhibitor (N34S) and the cystic fibrosis gene. *Am J Gastroenterol* 103(2): 368-374, 2008. PMID: 18076731.
- 27. Felderbauer P, Karakas E, Fendrich V, Bulut K, Werner I, Dekomien G, et al. Pancreatitis in primary hyperparathyroidism-related hypercalcaemia is not associated with mutations in the CASR gene. *Exp Clin Endocrinol Diabetes* 115(8): 527-529, 2007. PMID: 17853337.
- 28. Fishman D, Faulds G, Jeffery R, Mohamed-Ali V, Yudkin JS, Humphries S, et al. The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. *J Clin Invest* 102(7): 1369-1376, 1998. PMID: 9769329.
- Fjeld K, Weiss FU, Lasher D, Rosendahl J, Chen JM, Johansson BB, et al. A recombined allele of the lipase gene CEL and its pseudogene CELP confers susceptibility to chronic pancreatitis. *Nat Genet* 47(5): 518-522, 2015. <u>PMID: 25774637.</u>
- 30. Foley TP, Jr., Harrison HC, Arnaud CD and Harrison HE. Familial benign hypercalcemia. *J Pediatr* 81(6): 1060-1067, 1972. PMID: 4643023.
- Gao HK, Zhou ZG, Li Y and Chen YQ. Toll-like receptor 4 Asp299Gly polymorphism is associated with an increased risk of pancreatic necrotic infection in acute pancreatitis: a study in the Chinese population. *Pancreas* 34(3): 295-298, 2007. <u>PMID: 17414051.</u>
- 32. Gomes KB, Pardini VC and Fernandes AP. Clinical and molecular aspects of Berardinelli-Seip Congenital Lipodystrophy (BSCL). *Clin Chim Acta* 402(1-2): 1-6, 2009. <u>PMID: 19167372.</u>

- 33. Guenther A, Aghdassi A, Muddana V, Rau B, Schulz HU, Mayerle J, et al. Toll-like receptor 4 polymorphisms in German and US patients are not associated with occurrence or severity of acute pancreatitis. *Gut* 59(8): 1154-1155, 2010. <u>PMID: 20587548.</u>
- 34. Hofner P, Balog A, Gyulai Z, Farkas G, Rakonczay Z, Takacs T, et al. Polymorphism in the IL-8 gene, but not in the TLR4 gene, increases the severity of acute pancreatitis. *Pancreatology* 6(6): 542-548, 2006. <u>PMID:</u> <u>17124436</u>.
- 35. Kanth W, Reddy, DN. Genetics of acute and chronic pancreatitis: an update. *World Journal of Gastrointestinal Pathophysiology* 5(4): 427-437, 2014. PMID: 25400986.
- 36. Kawai T and Akira S. Signaling to NF-kappaB by Toll-like receptors. *Trends Mol Med* 13(11): 460-469, 2007. PMID: 18029230.
- 37. Kawai T and Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity* 34(5): 637-650, 2011. PMID: 21616434.
- 38. Kersten S. Physiological regulation of lipoprotein lipase. *Biochim Biophys Acta* 1841(7): 919-933, 2014. PMID: 24721265.
- 39. Kota SK, Krishna SV, Lakhtakia S and Modi KD. Metabolic pancreatitis: Etiopathogenesis and management. *Indian J Endocrinol Metab* 17(5): 799-805, 2013. <u>PMID: 24083160.</u>
- 40. Kumar S, Ooi CY, Werlin S, Abu-El-Haija M, Barth B, Bellin MD, et al. Risk Factors Associated With Pediatric Acute Recurrent and Chronic Pancreatitis: Lessons From INSPPIRE. *JAMA Pediatr* 170(6): 562-569, 2016. <u>PMID: 27064572.</u>
- 41. Lankisch PG, Blum T, Maisonneuve P and Lowenfels AB. Severe acute pancreatitis: when to be concerned? *Pancreatology* 3(2): 102-110, 2003. <u>PMID: 12748418.</u>
- 42. LaRusch J and Whitcomb DC. Genetics of pancreatitis. *Curr Opin Gastroenterol* 27(5): 467-474, 2011. PMID: 21844754.
- 43. Lerch MM, Mayerle J, Mahajan U, Sendler M, Weiss FU, Aghdassi A, et al. Development of Pancreatic Cancer: Targets for Early Detection and Treatment. *Dig Dis* 34(5): 525-531. <u>PMID: 27332960.</u>
- 44. Ma MH, Bai HX, Park AJ, Latif SU, Mistry PK, Pashankar D, et al. Risk factors associated with biliary pancreatitis in children. *J Pediatr Gastroenterol Nutr* 54(5): 651-656, 2011. <u>PMID: 22002481.</u>
- Magre J, Delepine M, Khallouf E, Gedde-Dahl T, Jr., Van Maldergem L, Sobel E, et al. Identification of the gene altered in Berardinelli-Seip congenital lipodystrophy on chromosome 11q13. *Nat Genet* 28(4): 365-370, 2001. <u>PMID: 11479539.</u>
- 46. Mahurkar S, Idris MM, Reddy DN, Bhaskar S, Rao GV, Thomas V, et al. Association of cathepsin B gene polymorphisms with tropical calcific pancreatitis. *Gut* 55(9): 1270-1275, 2006. <u>PMID: 16492714</u>.
- 47. Mansfield JC, Holden H, Tarlow JK, Di Giovine FS, McDowell TL, Wilson AG, et al. Novel genetic association between ulcerative colitis and the anti-inflammatory cytokine interleukin-1 receptor antagonist. *Gastroenterology* 106(3): 637-642, 1994. <u>PMID: 8119534</u>.
- 48. Merkel M, Eckel RH and Goldberg IJ. Lipoprotein lipase: genetics, lipid uptake, and regulation. *J Lipid Res* 43(12): 1997-2006, 2002. PMID: 12454259.
- 49. Morinville VD, Barmada MM and Lowe ME. Increasing incidence of acute pancreatitis at an American pediatric tertiary care center: is greater awareness among physicians responsible? *Pancreas* 39(1): 5-8, 2009. PMID: 19752770.
- Muddana V, Lamb J, Greer JB, Elinoff B, Hawes RH, Cotton PB, et al. Association between calcium sensing receptor gene polymorphisms and chronic pancreatitis in a US population: role of serine protease inhibitor Kazal 1type and alcohol. *World J Gastroenterol* 14(28): 4486-4491, 2008. <u>PMID: 18680227.</u>
- 51. Nauck M, Winkelmann BR, Hoffmann MM, Bohm BO, Wieland H and Marz W. The interleukin-6 G(-174)C promoter polymorphism in the LURIC cohort: no association with plasma interleukin-6, coronary artery disease, and myocardial infarction. *J Mol Med (Berl)* 80(8): 507-513, 2002. <u>PMID: 12185451.</u>
- 52. Neoptolemos JP, Raraty M, Finch M and Sutton R. Acute pancreatitis: the substantial human and financial costs. *Gut* 42(6): 886-891, 1998. PMID: 9691932.
- 53. Netea MG, Wijmenga C and O'Neill LA. Genetic variation in Toll-like receptors and disease susceptibility. *Nat Immunol* 13(6): 535-542, 2012. <u>PMID: 22610250.</u>
- 54. Nijmeijer RM, van Santvoort HC, Zhernakova A, Teller S, Scheiber JA, de Kovel CG, et al. Association analysis of genetic variants in the myosin IXB gene in acute pancreatitis. *PLoS One* 8(12): e85870, 2014. PMID: 24386489.
- Nydegger A, Heine RG, Ranuh R, Gegati-Levy R, Crameri J and Oliver MR. Changing incidence of acute pancreatitis: 10-year experience at the Royal Children's Hospital, Melbourne. J Gastroenterol Hepatol 22(8): 1313-1316, 2007. <u>PMID: 17489962.</u>
- 56. **O'Neill LA and Bowie AG**. The family of five: TIR-domain-containing adaptors in Toll-like receptor signalling. *Nat Rev Immunol* 7(5): 353-364, 2007. <u>PMID: 17457343.</u>

- 57. Ohyauchi M, Imatani A, Yonechi M, Asano N, Miura A, Iijima K, et al. The polymorphism interleukin 8 -251 A/T influences the susceptibility of Helicobacter pylori related gastric diseases in the Japanese population. *Gut* 54(3): 330-335, 2005. <u>PMID: 15710978.</u>
- 58. Pant C, Deshpande A, Olyaee M, Anderson MP, Bitar A, Steele MI, et al. Epidemiology of acute pancreatitis in hospitalized children in the United States from 2000-2009. *PLoS One* 9(5): e95552, 2014. PMID: 24805879.
- 59. Pant C, Deshpande A, Sferra TJ, Gilroy R and Olyaee M. Emergency department visits for acute pancreatitis in children: results from the Nationwide Emergency Department Sample 2006-2011. *J Investig Med* 63(4): 646-648. PMID: 25654293.
- Pearce SH, Wooding C, Davies M, Tollefsen SE, Whyte MP and Thakker RV. Calcium-sensing receptor mutations in familial hypocalciuric hypercalcaemia with recurrent pancreatitis. *Clin Endocrinol (Oxf)* 45(6): 675-680, 1996. PMID: 9039332.
- 61. Pollak MR, Brown EM, Chou YH, Hebert SC, Marx SJ, Steinmann B, et al. Mutations in the human Ca²⁺sensing receptor gene cause familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. *Cell* 75(7): 1297-1303, 1993. PMID: 7916660.
- Powell JJ, Fearon KC, Siriwardena AK and Ross JA. Evidence against a role for polymorphisms at tumor necrosis factor, interleukin-1 and interleukin-1 receptor antagonist gene loci in the regulation of disease severity in acute pancreatitis. *Surgery* 129(5): 633-640, 2001. <u>PMID: 11331456.</u>
- Rahalkar AR, Giffen F, Har B, Ho J, Morrison KM, Hill J, et al. Novel LPL mutations associated with lipoprotein lipase deficiency: two case reports and a literature review. Can J Physiol Pharmacol 87(3): 151-160, 2009. <u>PMID: 19295657.</u>
- 64. Restrepo R, Hagerott HE, Kulkarni S, Yasrebi M and Lee EY. Acute Pancreatitis in Pediatric Patients: Demographics, Etiology, and Diagnostic Imaging. *Am J Roentgenol* 206(3): 632-644, 2016. PMID: 26901022.
- Rodrigues R, Artieda M, Tejedor D, Martinez A, Konstantinova P, Petry H, et al. Pathogenic classification of LPL gene variants reported to be associated with LPL deficiency. J Clin Lipidol 10(2): 394-409, 2016. <u>PMID: 27055971.</u>
- Rosendahl J, Witt H, Szmola R, Bhatia E, Ozsvari B, Landt O, et al. Chymotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis. *Nat Genet* 40(1): 78-82, 2008. <u>PMID:</u> <u>18059268.</u>
- 67. Sargen K, Demaine AG and Kingsnorth AN. Cytokine gene polymorphisms in acute pancreatitis. *JOP* 1(2): 24-35, 2000. PMID: 11852287.
- 68. Schroder NW and Schumann RR. Single nucleotide polymorphisms of Toll-like receptors and susceptibility to infectious disease. *Lancet Infect Dis* 5(3): 156-164, 2005. <u>PMID: 15766650.</u>
- Sharer N, Schwarz M, Malone G, Howarth A, Painter J, Super M, et al. Mutations of the cystic fibrosis gene in patients with chronic pancreatitis. N Engl J Med 339(10): 645-652, 1998. <u>PMID: 9725921.</u>
- 70. Smithies AM, Sargen K, Demaine AG and Kingsnorth AN. Investigation of the interleukin 1 gene cluster and its association with acute pancreatitis. *Pancreas* 20(3): 234-240, 2000. <u>PMID: 10766448.</u>
- Takagi Y, Masamune A, Kume K, Satoh A, Kikuta K, Watanabe T, et al. Microsatellite polymorphism in intron 2 of human Toll-like receptor 2 gene is associated with susceptibility to acute pancreatitis in Japan. *Hum Immunol* 70(3): 200-204, 2009. <u>PMID: 19280717.</u>
- 72. Tukiainen E, Kylanpaa ML, Puolakkainen P, Kemppainen E, Halonen K, Orpana A, et al. Polymorphisms of the TNF, CD14, and HSPA1B genes in patients with acute alcohol-induced pancreatitis. *Pancreas* 37(1): 56-61, 2008. <u>PMID: 18580445.</u>
- 73. Vue PM, McFann K and Narkewicz MR. Genetic Mutations in Pediatric Pancreatitis. *Pancreas* 45(7): 992-996, 2015. <u>PMID: 26692446.</u>
- 74. Weiss FU, Schurmann C, Guenther A, Ernst F, Teumer A, Mayerle J, et al. Fucosyltransferase 2 (FUT2) non-secretor status and blood group B are associated with elevated serum lipase activity in asymptomatic subjects, and an increased risk for chronic pancreatitis: a genetic association study. *Gut* 64(4): 646-656. PMID: 25028398.
- 75. Whitcomb DC, Gorry MC, Preston RA, Furey W, Sossenheimer MJ, Ulrich CD, et al. Hereditary pancreatitis is caused by a mutation in the cationic trypsinogen gene. *Nat Genet* 14(2): 141-145, 1996. <u>PMID:</u> 8841182.
- Whitcomb DC, LaRusch J, Krasinskas AM, Klei L, Smith JP, Brand RE, et al. Common genetic variants in the CLDN2 and PRSS1-PRSS2 loci alter risk for alcohol-related and sporadic pancreatitis. *Nat Genet* 44(12): 1349-1354, 2012. <u>PMID: 23143602.</u>
- 77. Wilson AG, Symons JA, McDowell TL, McDevitt HO and Duff GW. Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. *Proc Natl Acad Sci U S A* 94(7): 3195-3199, 1997. <u>PMID: 9096369</u>.

- 78. Witt H, Beer S, Rosendahl J, Chen JM, Chandak GR, Masamune A, et al. Variants in CPA1 are strongly associated with early onset chronic pancreatitis. *Nat Genet* 45(10): 1216-1220, 2013. <u>PMID: 23955596</u>.
- 79. Witt H, Luck W, Hennies HC, Classen M, Kage A, Lass U, et al. Mutations in the gene encoding the serine protease inhibitor, Kazal type 1 are associated with chronic pancreatitis. *Nat Genet* 25(2): 213-216, 2000. PMID: 10835640.
- 80. Yang Z, Qi X, Wu Q, Li A, Xu P and Fan D. Lack of association between TNF-alpha gene promoter polymorphisms and pancreatitis: a meta-analysis. *Gene* 503(2): 229-234, 2012. <u>PMID: 22579868.</u>
- Yin YW, Sun QQ, Feng JQ, Hu AM, Liu HL and Wang Q. Influence of interleukin gene polymorphisms on development of acute pancreatitis: a systematic review and meta-analysis. *Mol Biol Rep* 40(10): 5931-5941, 2013. <u>PMID: 24072654.</u>
- Ying HY, Yu BW, Yang Z, Yang SS, Bo LH, Shan XY, et al. Interleukin-1B 31 C>T polymorphism combined with Helicobacter pylori-modified gastric cancer susceptibility: evidence from 37 studies. *J Cell Mol Med* 20(3): 526-536, 2016. <u>PMID: 26805397.</u>
- Zenker M, Mayerle J, Lerch MM, Tagariello A, Zerres K, Durie PR, et al. Deficiency of UBR1, a ubiquitin ligase of the N-end rule pathway, causes pancreatic dysfunction, malformations and mental retardation (Johanson-Blizzard syndrome). Nat Genet 37(12): 1345-1350, 2005. <u>PMID: 16311597.</u>
- 84. Zhang D, Zheng H, Zhou Y, Yu B and Li J. TLR and MBL gene polymorphisms in severe acute pancreatitis. *Mol Diagn Ther* 12(1): 45-50, 2008. <u>PMID: 18288881.</u>
- 85. Zhang Y, Liu C, Peng H, Zhang J and Feng Q. IL1 receptor antagonist gene IL1-RN variable number of tandem repeats polymorphism and cancer risk: a literature review and meta-analysis. *PLoS One* 7(9): e46017, 2012. <u>PMID: 23049925.</u>
- 86. **Zhou XJ, Cui Y, Cai LY, Xiang JY and Zhang Y**. Toll-like receptor 4 polymorphisms to determine acute pancreatitis susceptibility and severity: a meta-analysis. *World J Gastroenterol* 20(21): 6666-6670, 2014. <u>PMID: 24914392.</u>